Vegetable protein sources for carnivorous fish: potential and challenges

S. Refstie and T. Storebakken

Institute of Aquaculture Research, 6600 Sunndalsøra, Norway

stale.refstie@akvaforsk.nlh.no

Summary

In this review we describe how carnivorous fishes respond to vegetable protein feedstuffs. The amino acid profiles of gluten and soy products are complementary with respect to amino acid profile, and hold promise for further development of processing to increase protein contents and improve nutritional qualities. Soybeans are rich in antinutritional factors that disturb the digestion and/or physiology of carnivorous fish. Among these are heat-stable factor(s) inducing enteritis. Most soy antinutritional factors are removed by the thermal treatments followed by ethanol washing used to produce protein concentrates. Gluten products contain few antinutritional factors. As with most vegetable protein feedstuffs, gluten and soy protein concentrates do, however, contain phytic acid. Phytic acid-bound phosphorus is unavailable to fish, and phytic acid also binds essential divalent mineral elements, rendering them unavailable. The reduced availability of minerals has possible deleterious consequences and phytic acid should be eliminated by enzymic hydrolysis before feeding diets with high levels of plant protein to carnivorous fishes.

Keywords: carnivore, fish, feedstuffs, vegetable protein, antinutritional factors, processing

Introduction

Low temperature and steam dried fish meals are the commonly used protein sources in feeds for carnivorous fish. They are of high nutritional value, but are high-priced, and the supply is limited. Consequently there are major efforts to define and develop cost-effective protein sources that can, at least in part, be substitutes for fish meals in least-cost feed formulations. Problems related to bovine spongiform encephalopathy (BSE) have led restrictions on the use of animal by products in fish feeds. Thus, the main focus is on vegetable protein.

Legume seeds, such as peas, lupins, and soybeans (Hughes 1991; Watanabe *et al.* 1992; Shimeno *et al.*

1993; Kaushik et al. 1995; Olli et al. 1995; Robaina et al. 1995; Burel et al. 1998, 2000a), and lowglucosinolate rapeseeds (Hardy and Sullivan 1983; Burel et al. 2000a) have been used with success as partial substitutes for fish meal. Feeds for economically important cultured carnivorous fishes are, however, energy dense, and typically contain more than 40% protein, and at least 15% lipid. Because these fish digest starch and/or assimilate and metabolise glucose poorly (reviewed by Wilson 1994), the starch content of the diet is kept low. This calls for vegetable protein feedstuffs with high contents of protein and low contents of starch, sugars, and indigestible non-starch polysaccharides (NSP). Few seed crops meet these criteria, as legumes and oilseeds typically contain 25 to 40% protein and more than 30% carbohydrates. The seed commodity that comes closest is hulled and defatted (hexane extracted) soybean meal, but even this product is unsatisfactorily low in protein and high in NSP and sugars (Table 1).

Thus, the most feasible vegetable protein feedstuffs are the industrially manufactured protein concentrates. Corn gluten is already extensively used, and wheat gluten is being introduced. The soy industry has industrialised several processes that concentrate protein (reviewed by Lusas and Riaz 1995), and ethanol washed soy protein concentrates produced from hulled and defatted soy are of high nutritional quality for carnivorous fishes (Storebakken et al. 1998a; Berge et al. 1999; Mambrini et al. 1999; Kissil et al. 2000). Crude and refined soy products are reviewed in detail by Storebakken et al. (2000b). Extraction procedures to concentrate rapeseed protein are also developed (Jones 1979) but, although the resulting rapeseed concentrate is of high nutritional value (Higgs et al. 1994), this process is still not economically viable. Hulling and grinding followed by an air separation process may to some extent concentrate protein from legume seeds with low oil content (Booth et al. 2001).

Even when concentrated, the use of plant protein in feeds for carnivorous fish introduces several challenges. Compared with fish meal, the amino acid composition of vegetable protein is unbalanced, and excessive heating during industrial drying may reduce the protein quality of vegetable feedstuffs even further. More troublesome still is the fact that plants contain various antinutritional factors that disturb the digestion and/or physiology of fish. Carnivorous fishes have short intestines with little microflora (reviewed by Buddington *et al.* 1997), and therefore are very sensitive to such factors.

Protein quality

Compared to animal protein, plant protein is generally low in the indispensable amino acids arginine, lysine and methionine. For carnivorous fishes, the reported requirements range from 3 to 6% of crude protein (CP; N x 6.25) for arginine, 4 to 5% of CP for lysine, and 2 to 4% of CP for methionine (National Research Council 1993). As shown in Table 2, these requirements are well met by fish meal, but not by soy protein nor by gluten. The amino acid profiles of corn gluten and soy proteins are, however, complementary. Being a legume, soybean contains high levels of arginine and lysine in its protein, but little methionine. Corn gluten, on the other hand, is deficient in arginine and lysine, but has high methionine content. When combined, these vegetable protein feedstuffs produce a reasonably good

partial substitute for fish meal (Watanabe *et al.* 1993; Akiyama *et al.* 1995; Yamamoto *et al.* 1995).

Most commercial vegetable protein meals are residues from industrial manufacture of vegetable oil or starch. Such industrial processes usually include repeated heating to soften the seeds, remove solvents, and/or dry the meal residue. Excessive heating reduces the protein quality by amino acid cross—linking, binding to other nutrients, and/or oxidation (Finley and Phillips 1988; Davidek et al. 1990). This destroys the amino acids involved, and reduces the general digestibility of the protein (Ljøkjel et al. 2000). The most vulnerable amino acids are lysine (Maillard reaction) and cysteine (formation of disulfide bonds). When selecting industrially manufactured feedstuffs, it is important to be aware of and consider potential heat damage of the protein. Heat damage is avoided or reduced by moderate, short and moist heating.

Vegetable antinutritional factors

Vegetable antinutritional factors may broadly be divided into heat–stable and heat–labile factors. Unless these components can be inactivated or removed, they constitute the major restriction on the use of vegetable protein in fish feeds.

Table 1	Typical composition of fish meal and vegetable protein ingredients currently used in
	commercial feeds for carnivorous salmonid fish (percent of dry matter).

Protein source	Protein	Oil	Starch	NSP	Sugars
Fish meal ^{1,2}	78	12	_	_	_
Full fat soy ^{3,4}	42	21	3	18	11
Hulled and defatted soy3,4	57	1	3	23	14
Soy protein concentrate ^{3,4}	68	1	7	19	2
Corn gluten ^{2,4}	67	2	21	3	1
Wheat gluten ⁵	85	6	7	_	_

¹Anderson *et al.* 1993; ²Anderson *et al.* 1992; ³Lusas and Riaz 1995; ⁴Bach–Knudsen 1997: ⁵Storebakken *et al.* 2000a

Table 2 Typical critical indispensable (essential) amino acids in protein ingredients currently used in commercial feeds for salmonid fishes as compared to fish meal (g/16 g N = percent of CP).

	Fish meal ¹	Soybean ¹	Corn gluten ²	Wheat gluten ³
Arginine	5.9	7.3	3.7	3.6
Lysine	8.1	6.1	1.8	1.5
Methionine	3.0	1.4	2.3	1.6

¹Ljøkjel *et al.* 2000; ²Anderson *et al.* 1992; ³Storebakken *et al.* 2000a

As the term implies, heat-labile factors may be destroyed or inactivated by thermal processing. Glutens are practically free from such components, but legumes, and particularly soybeans, are rich in heat-labile antinutritional factors. The most significant of these are proteinase inhibitors and agglutinating lectins. Proteinase inhibitors are proteins capable of binding protein-hydrolysing digestive enzymes, thus restricting digestion and utilisation of dietary protein. Lectins are glycoproteins that bind (agglutinate) to receptors in the epithelium of the fish intestine (Hendriks et al. 1990), possibly with deleterious effects. Proteinase inhibitoractivity is commonly measured as mg bovine trypsin inhibited per g sample (TI-activity; Hammerstrand et al. 1981). Carnivorous fishes are sensitive to proteinase inhibitors (Krogdahl and Holm 1983; Takii et al. 1998), and both nutrient digestibility and growth by carnivorous fish are severely reduced if the dietary TI-activity exceeds 5 mg/g (Wilson and Poe 1985; Krogdahl et al. 1994; Olli et al. 1994). TI-activity may exceed 30 mg/g in raw soybeans. Certain beans, like navy or kidney beans, may also have a high TI-activity, whereas it is negligible in lupins and starch-rich legumes like peas and lentils (Table 3).

The most significant heat-stabile antinutritional factors in current vegetable protein feedstuffs for fish are antigens and phytic acid. Indigestible carbohydrates may also be troublemakers, and in particular the NSP.

Antigenicity is so far only investigated in salmonid fish, and only for soy products and wheat gluten. Soybeans contain antigenic factor(s) that induce enteritis in the distal intestine of salmonid fishes. This

inflammation is characterised by widening and shortening of the mucosal foldings, loss of the supranuclear vacuolisation of the absorptive cells in the intestinal epithelium, widening of and increased amounts of connective tissue in the central stroma within the mucosal foldings, and infiltration of a mixed leukocyte population in the lamina propria and submucosa (Baeverfjord and Krogdahl 1996). It is, furthermore, reflected in elevated lysozyme and IgM levels in the mucosa (Krogdahl et al. 2000), an increased number of circulating leukocyte cells, elevated circulating concentrations of plasma proteins and immunoglobulins, and increased activity of circulating neutrophils, monocytes, and macrophages (Rumsey et al. 1994). The condition is associated with a reduced absorptive capacity for nutrients by the distal intestine (Nordrum et al. 2000). How much the condition actually contributes to the reduced absorption of nutrients seen in soy-fed salmonids (see Figure 1) is unclear, as the distal intestine is not recognised as a major absorptive site in fish (Buddington et al. 1997). Furthermore, salmonid fish suffering the condition appear to grow normally (Refstie et al. 2000, 2001). On the other hand, the large intestine of fish absorbs macromolecules throughout life, and has important enteric immune functions (Buddington et al. 1997). The antigen(s) inducing this inflammatory response are still not identified. It is also unclear whether other plant seeds have similar antigenic properties, and whether the antigens(s) affect fish in general. The antigen(s) are apparently soluble in alcohol, as alcohol washed soy protein concentrates do not induce enteritis, whereas

Table 3 Contents in raw and processed legumes of Kunitz trypsin inhibitor (TI), Bowman–Birk combined trypsin and chymotrypsin inhibitor (BB–TI), and agglutinating lectins, together with activities of functional TIs, lectins, and urease.

		Contents of			Activities of	
Protein source	Kunitz-TI mg/g	BB–TI mg/g	Lectins mg/g	TIs mg/g*	Lectins mg/g**	Urease pH rise***
Raw pea ¹				1.3–1.8		
Raw lentil ¹				1.6		
Raw sweet lupin ²				0.1		
Raw navy bean ³				11.5		
Raw kidney bean ³				5.4		
Raw soybean						
Conventional ^{1,4,5,6,7}	30.3	10.7	8.3	17–31	2.3	2.1
Kunitz inhibitor-free ^{4,5}	0.04	11.4	8.0	5.6		2.0
Lectin-free ⁴	28.4	13.0	<0.0002			
Toasted soybean meal ^{4,6,7,8,}	9			3–9	0.01-0.2	<0.2
Soy protein concentrate ^{6,8}				2–7		

^{*} mg bovine trypsin inhibited per g meal

^{**} mg lectins agglutinating to brush border vesicles of chicken per g meal

^{***} pH rise in phosphate buffer as urease acts upon urea to produce ammonia

Pisulewska and Pisulewski 2000; ²Booth *et al.* 2001; ³Dhurandhar and Chang 1990; ⁴Han *et al.* 1991; ⁵Douglas *et al.* 1999; ⁶Anderson and Wolf 1995; ⁷Maenz *et al.* 1999; ⁸Refstie *et al.* 1999; ⁹Waldroup *et al.* 1985

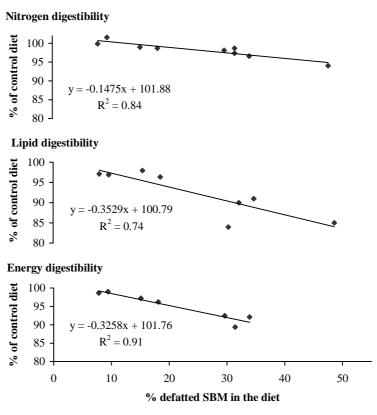


Figure 1 Reduced digestibility of nitrogen, lipid and energy by Atlantic salmon when replacing low temperature fish meal (LT–FM) by defatted and toasted soybean meal (SBM) in the diet. Fish oil was the only lipid source in all diets. Responses are presented relative to when feeding the LT–FM (control) diet. Data are taken from Refstie *et al.* 1998, 1999, 2000, 2001, and Storebakken *et al.* 1998b.

the alcohol extract (soy velasse) does (Ingh *et al.* 1996; Krogdahl *et al.* 2000). Wheat gluten, which may induce celiac sprue in humans, does not induce enteritis in salmon (Storebakken *et al.* 2000a).

Indigestible seed carbohydrates (Table 1) are largely in the form of soluble and insoluble NSP. Dietary soluble NSP may reduce lipid absorption by fish. For instance, indigestible starch (Grisdale–Helland and Helland 1997) and viscous guar gum (Storebakken 1985) restrict the absorption of protein and lipid by salmonids. Impaired lipid absorption when feeding soluble NSP is also a well–known and serious problem in chicken nutrition. Legumes and crucifers furthermore contain indigestible sugars (α–galactosides; Table 1) that produce diarrhoea in fish.

As shown in Table 4, the phosphorus content of fish meal is substantial. Thus, the concentration in diets for carnivorous fishes is high, and usually exceeds 10 g P/kg DM, but a major fraction of this is present in bone and thus is poorly available (Nordrum *et al.* 1997; Vielma *et al.* 1999). This results in a high P load to the aquatic environment, which may lead to algal blooms in freshwater systems (Vielma *et al.* 2000). Vegetable protein ingredients contain significantly less P than fish meal, and substitution of fish meal by vegetable

feedstuffs thus enables the use of less, and more highly digestible, P sources in the diet. As shown in Table 5 this is, however, complicated by the presence of phytic acid which is abundant in all types of vegetable protein feedstuffs that have a potential for use in feeds for carnivorous fish.

Phytic acid-bound P is unavailable to fish. Phytic acid also binds essential divalent mineral elements like Zn, and reduces the intestinal hydrolysis and thus the utilisation of dietary protein (Spinelli et al. 1983; Caldwell 1992). Lowered availability of Zn cause eye cataracts, as demonstrated in Chinook salmon (Oncorhynchus tshawytscha; Richardson et al. 1985). Baeverfjord et al. (1998) demonstrated that P deficiency in Atlantic salmon (Salmo salar) is initially manifested as a reduction in whole body Ca and P levels and the development of abnormally soft bones, although growth is little affected. In later stages, growth is severely impaired, and mortality increases. Reduced body Zn concentration is overcome by dietary supplementation of this element (Ramseyer et al. 1999); however, as indicated by Table 5, reduced body concentrations of Ca and P, and thus bone ash, seem harder to overcome.

It follows that vegetable antinutritional factors and nutrient classes from different feed ingredients may

Table 4 Typical phosphorus (P) and phytic acid in dry matter of fish meal and vegetable protein meals.

Ingredient	Fish meal ¹	Soybean ²	Pea ³	Rapeseed ³	Wheat gluten ¹
Phosphorus, g/kg	21.1	4.5	4.4	14.9	2.5
Phytic acid, g/kg	_	7.5	9.7	41.5	2.1
Phytic acid P as					
percent of total P	_	47	49	63	24

¹Storebakken *et al.* 2000a; ²Refstie *et al.* 1999; ³Burel *et al.* 2000b

Table 5 Availability of nitrogen and phosphorus in and elemental composition of Atlantic salmon grown for 84 days on diets with 60% fish meal or 15% fish meal and 48% untreated or phytase—treated soy protein concentrate as dietary protein sources. Soy protein concentrate diets were supplemented with dicalcium phosphate to obtain similar dietary P contents. From Storebakken *et al.* (1998a).

Dietary phytic acid, g/kg DM*	Fish meal diet 0.8	Soy protein concentrate diet 9.3	Phytase treated soy protein concentrate diet 0.5
Nitrogen			
Digestibility, %	91 ^a	85 ^b	88 ^{ab}
Retention, %	58 ^{ab}	55 ^b	62 ^a
Excretion, g/kg gain			
Faecal	4.8 ^b	8.2 ^a	5.9 ^b
Metabolic	17.0 ^a	16.3 ^a	13.8 ^b
Phosphorus			
Digestibility, %	41 ^b	30 ^c	49 ^a
Retention, %	30 ^a	25 ^b	32 ^a
Excretion, g/kg gain			
Faecal	8.1 ^a	8.7 ^a	6.3 ^b
Metabolic	1.4 ^a	0.5 ^b	1.9 ^a
Whole body concentration of			
Ash, g/kg	21.0 ^a	19.2 ^b	21.5 ^a
Ca, g/kg	4.2 ^a	3.5°	4.0 ^b
P, g/kg	4.4 ^a	4.0 ^b	4.4 ^a
Mg, mg/kg	403 ^a	389 ^b	413 ^a
Zn, mg/kg	61.6 ^a	42.6 ^b	60.6 ^a
Ratio Ca : P	0.94 ^a	0.85 ^b	0.91 ^a

 $^{^{\}mathrm{a,b,c}}$ Within rows, values not followed by the same superscript are significantly different (P< 0.05)

interact to affect the overall absorption of nutrients by fish. Although the contribution of each effect is small, the interactional sum may be significant. This is demonstrated in Figure 1, which shows how the digestibility of nutrients by Atlantic salmon is reduced by inclusion of toasted soybean meal in diets based on low temperature fish meal and fish oil. This appears to be a general response in carnivorous fishes, and inclusion of more than 20% soybean meal in the diet usually leads to reduced feed efficiency and slower growth (Watanabe *et al.* 1992; Shimeno *et al.* 1993; Kaushik *et al.* 1995; Olli *et al.* 1995).

From Table 3, it is apparent that raw soybean is the least acceptable protein feedstuff for carnivorous fishes. Defatted soy flakes are, however, typically toasted (steam-cooked) at 105°C for 30 minutes to remove solvent residues after the oil extraction procedure. This reduces the TI-activity down to levels tolerable by carnivorous fishes, and is accompanied by denaturation and inactivation of the lectins. Furthermore, feedstuffs in modern energy-dense fish feeds are subjected to a second moist heating during the high-pressure moist extrusion manufacturing of the diets. Thus, proteinase inhibitors and lectins are rarely

^{*} From dietary wheat

problematic when using defatted soy and/or other vegetable protein feedstuffs in modern fish feeds.

Heat-stable antinutritional factors are, however, hard to inactivate, and if not removed they restrict the use of plant proteins in fish feeds. Oligosaccharides, soluble NSP, and soy antigenic factor(s) that induce distal enteritis in salmonids (Ingh et al. 1996) are all soluble in alcohol, and so they are removed by ethanol and water washing in the manufacture of plant protein concentrates. This leaves phytic acid, which should be hydrolysed if vegetable feedstuffs provide a major part of the dietary protein. It is hydrolysed by microbial phytases, which are usually produced from Aspergillus niger. Current phytases are heat labile and consequently susceptible to inactivation during the high-pressure moist extrusion of feeds for carnivorous fish. Furthermore, phytase from A. niger has a temperature optimum of more than 50°C, and its hydrolytic activity is less than 10% of maximum at 10°C (Hoppe 1992). When used as a feed enzyme supplement, the phytase activity will be low at ambient water temperatures where cold-water carnivorous fishes are farmed. An alternative solution is pre-incubation of the vegetable protein sources with phytase, a procedure that is highly effective as shown in Table 5. If culturing fish at warm ambient water temperatures, post-extrusion application of liquid feed enzymes may prove a useful procedure to avoid thermal phytase destruction (Hughes and Soares 1998; Oliva-Teles et al. 1998, Vielma et al. 1998).

Future

Vegetable protein feedstuffs for carnivorous fishes need to have high protein contents and, in that respect, the most feasible candidates are gluten products. Soy protein concentrates are also promising, and such products are already available commercially. As the amino acid compositions of vegetable proteins are generally imbalanced for fish, it is important to combine vegetable protein feedstuffs with complimentary amino acid profiles. This is achieved by combining soy protein and corn gluten. It is also critical to eliminate and remove antinutritional factors. Soybeans are rich in antinutritional factors, but most of these are removed by industrial thermal treatments followed by ethanol washing to produce the commercial protein concentrates. Such manufacturing processes must be optimised to avoid protein damage by excessive heating. It is also desirable to eliminate NSP in vegetable feedstuffs in order to enhance the concentration of digestible energy in the feedstuffs. Finally, phytic acid should be eliminated to avoid disturbances of mineral element absorption and utilisation by fish fed high levels of plant protein. If and when these goals are achieved by cost-effective means, vegetable protein feedstuffs may replace substantial amounts of the fish meal currently used in energy-dense feeds for carnivorous fishes.

References

- Akiyama, T., Unuma, T., Yamamoto, T., Marcouli, P. and Kishi, S. (1995). Combination use of malt protein flour and soybean meal as alternative protein sources of fish meal in fingerling rainbow trout diets. *Fisheries Science* 61, 828–832.
- Anderson, R.L. and Wolf, W.R. (1995). Compositional changes in trypsin inhibitors, phytic acid, saponins and isoflavones related to soybean processing. *Journal of Nutrition* 125, 581S–588S.
- Anderson, J.S.., Lall, S.P., Anderson, D.M. and Chandrasoma, J. (1992). Apparent and true availability of amino acids from common feed ingredients for Atlantic salmon (*Salmo salar*) reared in sea water. *Aquaculture* 108, 111–124.
- Anderson, J.S., Lall, S.P., Anderson, D.M. and McNiven, M.A. (1993). Evaluation of protein quality in fish meals by chemical and biological assays. *Aquaculture* 115, 305–325.
- Bach–Knudsen, K.E. (1997). Carbohydrate and lignin contents of plant material used in animal feeding. *Animal Feed Science and Technology* 67, 319–338.
- Baeverfjord, G. and Krogdahl, Å. (1996). Development and regression of soybean meal induced enteritis in Atlantic salmon, *Salmo salar* L., distal intestine: a comparison with the intestines of fasted fish. *Journal of Fish Diseases* 19, 375–387.
- Baeverfjord, G., Åsgård, T. and Shearer, K.D. (1998). Development and detection of phosphorus deficiency in Atlantic salmon, *Salmo salar L.*, parr and post– smolts. *Aquaculture Nutrition* 4, 1–11.
- Berge, G.M., Grisdale–Helland, B. and Helland, S.J. (1999). Soy protein concentrate in diets for Atlantic halibut (*Hippoglossus hippoglossus*). *Aquaculture* 178, 139–148.
- Booth, M.A., Allan, G.L., Frances, J., and Parkinson, S. (2001). Replacement of fish meal in diets for Australian silver perch, *Bidyanus bidyanus* IV. Effects of dehulling and protein concentration on digestibility of grain legumes. *Aquaculture* 196, 67–85.
- Buddington, R.K., Krogdahl, Å. and Bakke–McKellep, A.M. (1997). The intestines of carnivorous fish: structure and functions and the relation with diet. *Acta Physiologica Scandinavica* 161, Suppl. 638, 67–80.
- Burel C., Boujard, T., Corraze, G., Kaushik, S.J., Boeuf, G., Mol, K.A., Geyten, S.v.d. and Kühn, E.R. (1998). Incorporation of high levels of extruded lupin in diets for rainbow trout (*Oncorhynchus mykiss*): nutritional value and effect on thyroid status. *Aquaculture* 163, 325–345.
- Burel C., Boujard, T., Kaushik, S.J., Boeuf, G., Geyten, S.v.d., Mol, K.A., Kühn, E.R., Quinsac, A., Krouti, M. and Ribaillier, D. (2000a). Potential plant–protein sources as fish meal substitutes in diets for turbot (*Psetta maxima*): growth, nutrient utilisation and thyroid status. *Aquaculture* 188, 363–382.
- Burel, C., Boujard, T., Tulli, F. and Kaushik, S.J. (2000b). Digestibility of extruded peas, extruded lupin, and

- rapeseed meal in rainbow trout (Oncorhynchus mykiss) and turbot (Psetta maxima). Aquaculture 188, 285-298
- Caldwell, R.A. (1992). Effect of calcium and phytic acid on the activation of trypsinogen and the stability of trypsin. Journal of Agricultural and Food Chemistry 40, 43-47.
- Davidek, J., Velisek, J. and Pokorny, J. (1990). Chemical Changes during Food Processing. Elsevier, Amsterdam, 448 pp.
- Dhurandhar, N.V. and Chang, K.C. (1990). Effect of cooking on firmness, trypsin inhibitors, lectins and cysteine/cysteine content of navy and red kidney beans (Phaseolus vulgaris). Journal of Food Science 55, 470-474.
- Douglas, M.W., Parsons, C.M. and Hymowitz, T. (1999). Nutritional evaluation of lectin-free soybeans for poultry. Poultry Science 78, 91-95.
- Finley, J.W. and Phillips, R.D. (1988). Protein Quality and Effects of Processing. Marcel Dekker, New York, NY, USA, 137 pp.
- Grisdale-Helland, B. and Helland, S.J. (1997). Replacement of protein by fat and carbohydrate in diets for Atlantic salmon (Salmo salar) at the end of the freshwater stage. Aquaculture 152, 167-180.
- Hammerstrand, G.E., Black, L.T. and Glover, J.D. (1981). Trypsin inhibitors in soy products: Modification of the standard analytical procedure. Cereal Chemistry 58, 42 - 45.
- Han, Y., Parsons, C.M. and Hymowitz, T. (1991). Nutritional evaluation of soybeans varying in trypsin inhibitor content. Poultry Science 70, 896-906.
- Hendriks, H.G.C.J.M., Ingh, T.S.G.A.M.v.d., Krogdahl, Å., Olli, J. and Koninkx, J.F.J.G. (1990). Binding of soybean agglutinin to small intestinal brush border membranes and brush border membrane enzyme activities in Atlantic salmon (Salmo salar). Aquaculture 91, 163-170.
- Higgs, D.A., Dosanjh, B.S., Prendergast, A.F., Beames, R.M., Hardy, R.W., Riley, W. and Deacon, G. (1994). Use of rapeseed/canola protein products in finfish diets. In: Nutrition and Utilization Technology in Aquaculture, pp.130-156 (eds. C. Lim, D.J. Seesa). AOAC Press, Champaign, USA.
- Hoppe, P.P. (1992). Review of the biological effects and the ecological importance of phytase in pigs. Presentations at the 4th BASF Animal Nutrition Forum, November 4–5, 1992. BASF, Ludwigshafen, Germany.
- Hughes, S.G. (1991). Use of lupin flour as a replacement for full-fat soy in diets for rainbow trout (Oncorhynchus mykiss). Aquaculture 93, 57-662.
- Hughes, K.P. and Soares, J.H. (1998). Efficacy of phytase on phosphorus utilization in practical diets fed to striped bass Morone saxatilis. Aquaculture Nutrition 4, 133-140.
- Ingh, T.S.G.A.M.v.d., Olli, J.J. and Krogdahl, Å. (1996). Alcohol-soluble components in soybeans cause morphological changes in the distal intestine of

- Atlantic salmon, Salmo salar L. Journal of Fish Diseases 19, 47-53.
- Jones, J.D. (1979). Rapeseed protein concentrate preparation and evaluation. Journal of the American Oil Chemists Society 56, 716-721.
- Kaushik, S.J., Cravedi, J.P., Lalles, J.P., Sumpter, J., Fauconneau, B. and Laroche, M. (1995). Partial or total replacement of fish meal by soybean protein on growth, protein utilization, potential estrogenic or antigenic effects, cholesterolemia and flesh quality in rainbow trout, Oncorhynchus mykiss. Aquaculture 133, 257-274.
- Kissil, G.W., Lupatsch, I., Higgs, D.A. and Hardy, R.W. (2000). Dietary substitution of soy and rapeseed protein concentrates for fish meal, and their effects om growth and nutrient utilization in gilthead seabream Sparus aurata L. Aquaculture Research 31, 595–601.
- Krogdahl, Å. and H.H. Holm (1983). Pancreatic proteinases from man, trout, pig, cow, chicken, mink and fox enzyme activities and inhibition by soybean and lima bean inhibitors. Comparative Biochemistry and Physiology 74B, 403-409.
- Krogdahl, Å., Lea, T.B. and Olli, J.J. (1994). Soybean proteinase inhibitors affect intestinal trypsin activities and amino acid digestibilities in rainbow trout (Oncorhynchus mykiss). Comparative Biochemistry and Physiology 107A, 215-219.
- Krogdahl, Å., Bakke-McKellep, A.M., Røed, K.H. and Baeverfjord, G. (2000). Feeding Atlantic salmon Salmo salar L. soybean products: effects on disease resistance (furunculosis), and lysozyme and IgM levels in the intestinal mucosa. Aquaculture Nutrition 6, 77-84.
- Ljøkjel, K., Harstad, O.M. and Skrede, A. (2000). Effect of heat treatment of soybean meal and fish meal on amino acid digestibility in mink and dairy cows. Animal Feed Science and Technology 84, 83–95.
- Lusas, E.W. and Riaz, M.N. (1995). Soy protein products: Processing and use. Journal of Nutrition 125, 573S-
- Maenz, D.D., Irish, G.G. and Classen, H.L. (1999). Carbohydrate-binding and agglutinating lectins in raw and processed soybean meals. Animal Feed Science and Technology 76, 335-343.
- Mambrini, M., Roem, A.J., Cravèdi, J.P., Lallès, J.P. and Kaushik, S.J. (1999). Effects of replacing fish meal with soy protein concentrate and of DL-methionine supplementation in high-energy, extruded diets on the growth and nutrient utilization of rainbow trout, Oncorhynchus mykiss. Journal of Animal Science 77, 2990-2999.
- National Research Council (1993). Nutrient Requirements of Fish. National Academy Press, Washington, USA, 114 pp.
- Nordrum, S., Åsgård, T., Shearer, K.D. and Arnessen, P. (1997). Availability of phosphorus in fish bone meal and inorganic salts to Atlantic salmon (Salmo salar) as determined by retention. Aquaculture 157, 51-61.
- Nordrum, S., Bakke-McKellep, A.M., Krogdahl, Å. and Buddington, R.K. (2000). Effects of soybean meal and

- salinity on intestinal transport of nutrients in Atlantic salmon (Salmo salar L.) and rainbow trout (Oncorhynchus mykiss). Comparative Biochemistry and Physiology 125B, 317-335.
- Olivia-Teles, A., Periera, J.P., Gouveia, A. and Gomes, E. (1998). Utilisation of diets supplemented with microbial phytase by seabass (Dicentrarchus labrax) juveniles. Aquatic Living Resources 11, 255-259.
- Olli, J.J., Hjelmeland, K. and Krogdahl, Å. (1994). Soybean trypsin inhibitors in diets for Atlantic salmon (Salmo salar, L): effects on nutrient digestibilities and trypsin in pyloric caeca homogenate and intestinal content. Comparative Biochemistry and Physiology 109A, 923-928.
- Olli, J.J., Krogdahl, Å. and Våbenø, A. (1995). Dehulled solvent-extracted soybean meal as a protein source in diets for Atlantic salmon, Salmo salar L. Aquaculture Research 26, 167-174.
- Pisulewska, E. and Pisulewski, P.M. (2000). Trypsin inhibitor activity of legume seeds (peas, chickling vetch, lentils, and soya beans) as affected by the technique of harvest. Animal Feed Science and Technology 86, 261-265.
- Ramseyer, L., Garling, D., Hill, G. and Link, J. (1999). Effect of dietary zinc supplementation and phytase pretreatment of soybean meal or corn gluten meal on growth, zinc status and zinc-related metabolism in rainbow trout (Oncorhynchus mykiss). Fish Physiology and Biochemistry 20, 251–261.
- Refstie, S., Storebakken, T. and Roem, A.J. (1998). Feed consumption and conversion in Atlantic salmon (Salmo salar) fed diets with fish meal, extracted soybean meal or soybean meal with reduced content of oligosaccharides, trypsin inhibitors, lectins and soya antigens. Aquaculture 162, 301-312.
- Refstie, S., Svihus, B., Shearer, K.D. and Storebakken, T. (1999). Non-starch polysaccharides in soybean meals and effects on the absorption of nutrients in farmed Atlantic salmon and broiler chickens. Animal Feed Science and Technology 79, 331-345.
- Refstie, S., Korsøen, Ø.J., Storebakken, T., Baeverfjord, G., Lein, I. and Roem, A.J. (2000). Differing nutritional responses to dietary defatted soybean meal in rainbow trout (Oncorhynchus mykiss) and Atlantic salmon (Salmo salar). Aquaculture 190, 49-63.
- Refstie, S., Storebakken, T., Baeverfjord, G. and Roem, A.J. (2001). Long-term protein and lipid growth of Atlantic salmon (Salmo salar) fed diets with partial replacement of fish meal by soy protein products at medium or high lipid level. Aquaculture 193, 91-106.
- Richardson, N.L., Higgs, D.A., Beames, R.M. and McBride, J.R. (1985). Influence of dietary calcium, phosphorus, zinc and sodium phytate level on cataract incidence, growth and histopathology in juvenile Chinook salmon (Oncorhynchus tsawytscha). Journal of Nutrition 115, 553-567.
- Robaina, L., Izquierdo, M.S., Moyano, F.J., Socorro, J., Vergara, J.M., Montero, D. and Fernández-Palacios, H. (1995). Soybean and lupin seed meals as protein sources in diets for gilthead seabream (Sparus aurata):

- nutritional and histological implications. Aquaculture 130, 219–233.
- Rumsey, G.L., Siwicki, A.K., Anderson, D.P. and Bowser, P.R. (1994). Effect of soybean protein on serological response, non-specific defence mechanisms, growth, and protein utilization in rainbow trout. Veterinary Immunology and Immunopathology 41, 323-339.
- Shimeno, S., Kumon, M., Ando, H. and Ukawa, M. (1993). The growth performance and body composition of young yellowtail fed with diets containing defatted soybean meals for a long period. Nippon Suisan Gakkaishi 59, 821-825.
- Spinelli, J., Houle, C.R. and Wekell, J.C. (1983). The effect of phytates on the growth of rainbow trout (Salmo gairdneri) fed purified diets containing varying quantities of calcium and magnesium. Aquaculture 30, 71-83.
- Storebakken, T. (1985). Binders in fish feeds I. Effect of alginate and guar gum on growth, digestibility, feed intake and passage through the gastrointestinal tract of rainbow trout. Aquaculture 47, 11-26.
- Storebakken, T., Shearer, K.D. and Roem, A.J. (1998a). Availability of protein, phosphorus and other elements in fish meal, soy-protein concentrate and phytasetreated soy-protein-concentrate-based diets to Atlantic salmon, Salmo salar. Aquaculture 161, 365-379.
- Storebakken, T., Kvien, I.S., Shearer, K.D., Grisdale-Helland, B., Helland, S.J. and Berge, G.M. (1998b). The apparent digestibility of diets containing fish meal, soybean meal or bacterial meal fed to Atlantic salmon (Salmo salar): evaluation of different faecal collection methods. Aquaculture 169, 195-210.
- Storebakken, T., Shearer, K.D., Baeverfjord, G., Nielsen, B.G., Åsgård, T., Scott, T. and De Laporte, A. (2000a). Digestibility of macronutrients, energy and amino acids, absorption of elements and absence of intestinal enteritis in Atlantic salmon, Salmo salar, fed diets with wheat gluten. Aquaculture 184, 115-132.
- Storebakken, T., Refstie, S. and Ruyter, B. (2000b). Soy products as fat and protein sources in fish feeds for intensive aquaculture. In: Drackley, J.K. (Ed.), Soy in Animal Nutrition, pp. 127-170 (ed. J.K. Drackley), Federation of Animal Science Societies, Savoy, IL, USA.
- Takii, K., Nakamura, M., Urakawa, K., Miyashita, S., Nasu, T., Kubo, Y., Tanaka, Y. and Kumai, H. (1998). Soybean trypsin inhibitors inhibit trypsin-like and basic proteinase activities of cultured fishes. Fish Science 64, 935-938.
- Vielma, J., Lall, S.P., Koskela, J., Schöner, F.-J. and Mattila, P. (1998). Effects of dietary phytase and cholecalciferol on phosphorus bioavailability in rainbow trout (Oncorhynchus mykiss). Aquaculture 163, 309-323.
- Vielma, J., Ruohonen, K. and Lall, S.P. (1999). Supplemental citric acid and particle size of fish bonemeal influence the availability of minerals in rainbow trout Oncorhynchus mykiss (Walbaum). Aquaculture Nutrition 5, 65–71.

- Vielma, J., Mäkinen, T., Ekholm, P. and Koskela, J. (2000). Influence of dietary soy and phytase levels on performance and body composition of large rainbow trout (*Oncorhynchus mykiss*) and algal availability of phosphorus load. *Aquaculture* 183, 349–362.
- Waldroup, P.W., Ramsey, B.E., Hellwig, H.M. and Smith, N.K. (1985). Optimum processing for soybean meal used in broiler diets. *Poultry Science* 64, 2314–2320.
- Watanabe, T., Viyakarn, V., Kimura, H., Ogawa, K., Okamoto, N. and Iso, N. (1992). Utilization of soybean meal as a protein source in a newly developed soft–dry pellet for yellowtail. *Nippon Suisan Gakkaishi* 58, 1761–1773.
- Watanabe, T., Pongmaneerat, J., Satoh, S. and Takeuchi, T. (1993). Replacement of fish meal by alternative protein sources in rainbow trout diets. *Nippon Suisan Gakkaishi* 59, 1573–1579.
- Wilson, R.P. (1994). Utilization of dietary carbohydrate by fish. *Aquaculture* 124, 67–80.
- Wilson, R.P. and Poe, W.E. (1985). Effect of feeding soybean meal with varying trypsin inhibitor activities on growth of fingerling channel catfish. *Aquaculture* 46, 19–25.
- Yamamoto, T., Unuma, T. and Akiyama, T. (1995). The effect of combined use of several alternative protein sources in fingerling rainbow trout diets. *Fisheries Science* 61, 915–920.